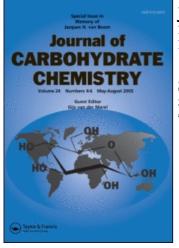
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Syntheses of D-*Myo***-Inositol-1,2,6-Trisphosphate and -2,6-bisphosphate** Sung-Kee Chung^a; Seok-Ho Yu^a; Young-Tae Chang^a

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SYNTHESES OF D-MYO-INOSITOL-1,2,6-TRISPHOSPHATE

AND -2,6-BISPHOSPHATE

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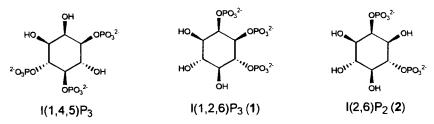
ABSTRACT

A D-myo-inositol derivative (3), obtained from methyl α -D-glucopyranoside by Ferrier rearrangement, was efficiently transformed to D-myo-inositol 1,2,6-trisphosphate (1, α -trinositol) and D-myo-inositol 2,6-bisphosphate (2).

INTRODUCTION

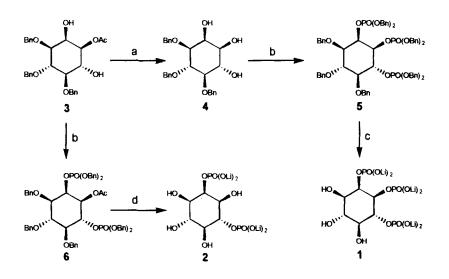
D-myo-Inositol 1,4,5-trisphosphate [D-I(1,4,5)P₃], released into the cytosol of cells by the phospholipase C (PLC)-catalyzed cleavage of phosphatidylinositol 4,5bisphosphate [PI(4,5)P₂], is known to mobilize intracellular calcium.¹ A number of other myo-inositol phosphates have also been implicated as either second messengers or key metabolic intermediates in the intracellular signal transduction pathways. D-myo-Inositol 1,2,6-trisphosphate (1, α -trinositol; PP56), obtained from InsP₆ (phytic acid) by enzymatic hydrolysis with yeast phytase, was shown to possess no appreciable agonist activity in biological systems, but to have an inhibitory effect on neuropeptide Y-evoked vasoconstriction in many vascular assays, and was also shown to be a potent antiinflammatory agent.² More recently it has been suggested that α -trinositol has a specific and possibly unique binding site in membranes from several rat tissues although it also shares common binding characteristics with Ins(1,3,4,5)P₄ and, to a lesser degree, also with InsP₆ and Ins(1,4,5)P₃.³ A couple of chemical⁴ and enzyme assisted⁵ syntheses of α -trinositol and its analogues⁶ were reported.

As a part of our efforts to understand the molecular recognition aspect of inositol phosphate-dependent signal transduction including phospholipase C, inositol 1,4,5-trisphosphate receptor, and the metabolic enzymes, we have been studying syntheses and biological activities of various natural and unnatural inositols and their phosphate analogues.⁷ Synthesis of modified *myo*-inositol derivatives might be carried out with *myo*-inositol as the starting material, but this approach suffers from the inherent disadvantage of having to resolve the racemates at a suitable stage of the synthesis, if an optically active compound is desired. Use of chiral natural products as the starting material has some advantage in this respect. Ferrier rearrangement⁸ has been sporadically employed for the conversion of a sugar derivatives into chiral inositol derivatives.⁹ We report here the application of the Ferrier methodology to the synthesis of D-*myo*-inositol 1,2,6-trisphosphate (1) and its 1-dephosphorylated analogue, D-*myo*-inositol 2,6-bisphosphate (2).



RESULTS AND DISCUSSION

1-O-Acetyl-3,4,5-tri-O-benzyl-D-myo-inositol (3) was prepared by the Ferrier rearrangement from D-glucose essentially according to the reported procedure.^{9a} Base catalyzed solvolysis of 3 using MeONa/MeOH gave D-myo-I(3,4,5)Bn₃ (4) in 95% yield. Compound 4 was phosphorylated by successive treatments with N,N-diisopropyldibenzylphosphoramidite and 1H-tetrazole, and then H₂O₂ to give protected



Scheme. a. NaOMe, MeOH. b. (i) *N*,*N*-diisopropyldibenzylphosphoramidite, 1H-tetrazole, DMF, (ii) 30% H_2O_2 . c. (i) H_2 (1 atm), Pd(OH)₂, MeOH. (ii) pH 10 (LiOH). d. H_2 , Pd(OH)₂. (ii) 1N LiOH, (iii) H⁺ ion exchange, (iv) pH 10 (LiOH).

I(1,2,6)P₃, 5. Hydrogenolysis of 5 in the presence of Pd(OH)₂ catalyst was followed by addition of LiOH to pH 10 to give the lithim salt of α -trinositol, 1.

Direct phosphorylations of 3 with the same procedure as described for 6 gave protected D-I(2,6)P₂. Hydrogenolysis and the base catalyzed hydrolysis of 6 followed by ion-exchange chromatography and pH adjustment to 10 gave the lithium salt of D-I(2,6)P₂.

Thus, we accomplished the chemical synthesis of biologically interesting natural product, α -trinositol and the first synthesis of D-myo-inositol 2,6-bisphosphate (2) which might be useful in evaluating the importance of the 2-phosphate group in the action of α -trinositol.

EXPERIMENTAL

3,4,5-Tri-O-benzyl-D-myo-inositol (4). A solution of 3 (80.8 mg, 0.16 mmol) and sodium methoxide (20.3 mg, 0.36 mmol) in anhydrous methanol (2 mL) was stirred at room temperature for 30 min. The mixture was poured into a saturated aq NaHCO₃

solution and extracted with CH₂Cl₂. The combined organic layer was dried over anhydrous Na₂SO₄ and concentrated to give a white solid 4 (70.2 mg, 95%). ¹H NMR (CDCl₃): δ 3.30 (t, J = 9.4, 1H, H-5), 3.38 (dd, J = 9.4, 3.1 Hz, 1H, H-1), 3.49 (dd, J = 9.4, 3.1 Hz, 1H, H-3), 3.87 (t, J = 9.4 Hz, 1H, H-6), 3.93 (t, J = 9.4 Hz, 1H, H-4), 4.18 (t, J = 3.1 Hz, 1H, H-2), 4.70-4.95 (m, 6H, 3PhCH₂), 7.29-7.32(m, 15H, Ph); ¹³C NMR (CDCl₃): δ 69.21, 71.83, 72.78, 72.84, 75.36, 75.76 (inositol ring carbons), 80.19, 81.22, 82.58 (3PhCH₂), 127.04-135.57 (Ph); Mass Spectrum (FAB), *m/z* 473 (M⁺+ 23); [α]_D³² -2.7 (*c* 1.79, CHCl₃).

3,4,5-Tri-O-benzyl-D-myo-inositol 1,2,6-tris(dibenzyl phosphate) (5). To a solution of 4 (46 mg, 0.102mmol) and 1H-tetrazol (28 mg, 0.40 mmol) in dimethylformamide (2 mL), was added N,N-diisopropyldibenzylphosphoramidite (352.6 mg, 1.02 mmol), and the mixture was stirred for 5 h at room temperature. Sodium phosphate buffer (1N, pH 7, 2 ml) and 30% ag H_2O_2 (3 mL) were added and the solution was stirred overnight. Ethyl acetate was added to the mixture, and the organic layer was separated and washed with 1% aq KHSO₄, 1% aq NaHCO₃, and then saturated aq NaCl. The organic layer was dried over anhydrous MgSO4, filtered and the filtrate was concentrated under reduced pressure. The residue was separated by silica gel chromatography to give 5 as an oil (76.8 mg, 65%). ¹H NMR (CDCl₃): δ 3.47 (dd, J = 9.3, 3.1 Hz, 1H, H-3), 3.49 (app.t, J = 9.3 Hz, 1H, H-5), 3.88 (app.t, J = 9.3 Hz, 1H, H-4), 4.34 (app.t, J = 8.7 Hz, 1H, H-1), 4.43-4.85 (m, 6H, 3PhCH₂), 4.88-5.19 (m, 13H, 6PhCH₂ & H-6), 5.48(br d, J = 8.7 Hz, 1H, H-2), 7.01-7.42 (m, 45H, Ph); ¹³C NMR (CDCl₃) & 69.09-69.90 [3P(O)(OCH₂Ph)₂], 72.71, 74.62, 74.69, 75.29, 75.81, 77.40, 78.24, 80.39, 80.67 (inositol ring carbons & 3PhCH₂), 138.8-127.96 (Ph); ³¹P NMR $(CDCl_3)$; δ -1.42, -0.46, -0.06; $[\alpha]_D^{29} = -2.1$ (c 2.5, CHCl₃). Crude 5 was used in the next step without further purification.

Lithium salt of α -trinositol (1). A mixture of 5 (63.7 mg, 55 µmol) and Pd(OH)₂ (100 mg) in methanol (5 mL) was stirred under H₂ gas (1 atm) at room temperature. After 3 days the mixture was filtered through celite and washed with water. The filtrate was concentrated under reduced pressure, and the residual aqueous solution was adjusted to pH 10 with 1N aqueous LiOH, and then lyopilized to give 1 as a white powder. ¹H NMR (D₂O, pH 10) δ 3.42 (dd, J = 10.0, 1.9 Hz, 1H, H-3), 3.48(app. t, J = 9.4 Hz, 1H,

H-5), 3.81 (app. t, J = 10.0 Hz, 1H, H-4), 3.92 (m, 1H, H-1), 4.23 (app. q, J = 8.7 Hz, 1H, H-6), 4.68 (br d, J = 6.8 Hz, 1H, H-2); ¹³C NMR (D₂O, pH 10): δ 74.64, 75.15, 75.96, 77.75, 78.12 (2C); ³¹P NMR (D₂O, pH 10) δ 5.08, 5.44, 5.86; $[\alpha]_D^{29} = -16.9$ (c 0.56, H₂O, pH 10), lit. ^{5b} $[\alpha]_D^{25}$ -19.5 (c 1.1, H₂O).

1-O-Acetyl-3,4,5-tri-O-benzyl-D-mvo-inositol 2,6-bis(dibenzyl phosphate) (6) To a solution of 3^{9a} (90 mg, 0.18 mmol) and 1H-tetrazol (50 mg) in CH₂Cl₂ (10 mL), was added N,N-diisopropyldibenzylphosphoramidite (330 mg, 0.95 mmol), and the mixture was stirred for 3 h at room temperature. Sodium phosphate buffer (1N, pH 7, 3 mL) and H_2O_2 (30%, 3 ml) were added and the solution was stirred overnight. Ethyl acetate was added to the mixture, and the organic layer was separated and washed with 1% aq KHSO₄, 1% aq NaHCO₃, and then saturated aq NaCl. The organic layer was dried over anhydrous MgSO₄, filtered and the filtrate was concentrated under reduced pressure. The product mixture was separated by silica gel chromatography to give 6 as an oil (87 mg, 48 %). ¹H NMR (CDCl₃) δ 1.78 (s, 3H, CH₃CO), 3.56-3.64 (m, 2H, H-3 & H-5), 3.92 (dd, J = 9.4, 9.3 Hz, 1H, H-4), 4.55-5.26 (m, 17H, 7PhCH₂, H-1, H-2 & H-6), 7.06-7.44 (m, 35H, Ph); 13 C NMR (CDCl₃) δ 20.52 (CH₃CO), 69.14–69.44 [2P(O)(OCH₂Ph)₂], 70.23, 74.18, 77.59, 78.34, 80.49, 81.12 (inositol ring carbons), 72.58, 75.41, 75.89 (3PhCH₂, assigned by DEPT-135), 170.26 (CH₃CO); ³¹P NMR (CDCl₃) δ -1.09, -0.61; $[\alpha]_{D}^{25} = +0.33$ (c 1.64, CHCl₃). Crude 6 was used in the next step without further purification.

Lithium salt of D-myo-inositol 2,6-bisphosphate (2). A mixture of 6 (32.7 mg, 32.3 μ mol) and Pd(OH)₂ (68 mg) in methanol (3 mL) was stirred under H₂ gas (1 atm) at room temperature. After 3 days the mixture was filtered through celite, and washed with water. 1N LiOH (3 mL) was added to the filtrate and the mixture was stirred at 80 °C. After 3 h the mixture was cooled to room temperature and then passed through the ion exchange resin (Dowex 50W-hydrogen, strongly acidic). The aqueous layer was adjusted to pH 10 with 1N aqueous LiOH and lyophilized to give compound 2. ¹H NMR (D₂O, pH 10) δ 3.45 (app t, J = 9.3 Hz, 1H, H-5), 3.49 (dd, J = 10.0, 2.5 Hz, 1H, H-3), 3.63 (br d, J = 9.3 Hz, 1H, H-1), 3.82 (app t, J = 10.0 Hz, 1H, H-4), 4.20 (app. q, J = 9.3 Hz, 1H, H-6), 4.52 (br d, J = 9.3 Hz, 1H, H-2); ³¹P NMR (D₂O, pH 10) δ 5.55, 5.61; [α]_D²² = - 4.3 (*c* 0.7, H₂O, pH 10). The spectral data of 2 were satisfactorily compared with those of DL-myo-inositol 2,6-bisphosphate which was prepared by a different synthetic route.¹⁰

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